Isolation of Some Enteropathogens from Retailed Poultry Meat in Alexandria Province

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ABSTRACT

A total of 100 random samples of chicken meat, chicken nuggets, chicken paneehh and chicken luncheon (25 of each) were collected from different supermarkets in Alexandria province. The collected samples were subjected to bacteriological examination for detection of enteropathogens. The obtained results as following; *Salmonella* isolated from chicken meat, chicken nuggets, chicken paneehh and chicken luncheon as 56, 8, 12 and 8 %, respectively. *E.coli* was isolated by 68, 12, 12 and 8 % in chicken meat, chicken nuggets, chicken paneehh and chicken luncheon, respectively. While, *Campylobacter jejuni* was detected in 76, 16 in chicken meat and chicken paneeh, respectively and it could not be isolated as 60, 8 and 90 % from chicken (meat, nuggets and luncheon), respectively but could not be isolated from chicken nuggets and luncheon. *Aeromonashydrophila* isolated at percentages of 28 and 4 % in chicken meat and chicken paneehh and chicken paneehh and chicken nuggets and luncheon.

INTRODUCTION

Poultry meat and its products are very popular food in Egypt as well as throughout the world. No wonder since it is delicious, nutritious and considered as a good and cheap source of protein characterized by good flavour and easily digested.

Escherichia coli, Campylobacter and *Salmonella* are often present on fresh tissues because the slaughtering process does not include bacterial steps. Since poultry is a major food source of Salmonella, its contamination may result in the development of human illness. Also, Campylobacter spp. has been recognized as one of the most common cause diarrhea or enterocolitis (Lee et al., 1998).

Cebedo et al., (2008) concluded that Salmonella are pathogenic bacteria that can contaminate food products during or after processing. Ready-to-eat food dose not undergo any treatment to ensure its safety before consumption, and therefore risk of foodborne diseases must be considered if these pathogens are present in food.

Aruno et al., (2007) found that *E.coli O157: H* 7 is one of the major threats to public health due to consumption of insufficient cooked meat and meat products. The microorganism is known as a foodborne pathogens evening the presence of low levels.

Shriver-lake et al., (2007) mentioned that food poisoning causes untold discomfort to many people each year. One of the primary culprits in food poisoning is *E.coli O157:H7*, while most cases cause intestinal discomfort, up to 7 % of the incidence leads to a severe complication called hemolytic uremic syndrome which may be fatal.

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Bacillus cereus causes two different types of foodborne illness, the diarrheal type and emetic type. The diarrheal type of foodborne illness is caused by enterotoxins produced during vegetative growth of Bacillus cereus in the small intestine, whereas the emetic toxic is produced by cells growing in the food. For both types of foodborne illness, the food involved has usually been treated and surviving spores are the sources of food poisoning. Although Bacillus cereus is not a competitive microorganism, it grows well after cooking and cooling (< 48 °C). Heat treatments causes spore germination and in the absence of competing flora Bacillus cereus grow well (Granum, 2001).

Aeromonas hydrophila has been found in red meat and poultry. The elaboration of toxin by the microorganism at low temperatures may have significance in foodborne disease when ingested as performed toxin in food (**Biair et al.**, **2000**).

From the previous information, the purpose of this study is to give an available assay about isolation of some enteropathogens from retailed poultry meat in Alexandria province. These microbes such as Salmonella, E.coli, Campylobacter jejuni, Bacillus cereus and Aeromonas hydrophila.

MATERIAL AND METHODS

A total of 100 samples from chicken meat, chicken nuggets, chicken paneeh and chicken luncheon (25 from each) were collected from different supermarkets in Alexandria province and transferred as quickly as possible and rapidly to the lab to avoid any change in the quality of sample due to microbial action.

Preparation of samples for bacteriological examination (ICMSF, 1978):

Chicken meat and chicken luncheon samples were firstly cauterized by using hot spatula (surface sterilization) then the cauterized parts were removed by using sterilized scalpel and forceps, while, the chicken paneeh and chicken nuggets samples were firstly thawed by holding in refrigerator at 3-4°C for 1 hour. Then under complete aseptic conditions 10 grams of each sample were weighed and transferred into a sterile homogenizer flask containing 90 ml of sterile peptone water 0.1 %. The contents of the homogenizer flask were homogenized for 2.5 minutes at room temperature. One ml of the original dilution (10⁻¹) was transferred to 9 ml of sterile peptone water (0.1%) to prepare further decimal dilutions up to 10^{-6} .

- Isolation of Salmonella according to (APHA, 1992).
 Sterile selenite F broth (Gibco) and Brilliant green agar (Biolife)
- 2- Isolation of *E.coli* according to (ICMSF, 1978). MacConkey's broth (Adwic), Brilliant green bile 2 % (Oxoid) and Eosine methylene blue agar (Biolife).
 3- Isolation of *Campylobacter jejuni*
- 3- Isolation of Campylobacter jejuni according to (Corry et al., 2001): Bolton broth (Oxoid) and Charcoal cefoperazone deoxycholate agar (CCDA) (Oxoid).
- 4- Isolation of Bacillus cereus according to (APHA, 1992).
 Polymexine pyruvate egg-yolk mannitol Bromothylmol blue agar (PEMPA) (Oxoid).
- 5- Isolation of Aeromonas hydrophila by using: Aeromonas base medium (Ryan's formulation) (Oxoid).

Incidence of Salmonella, E.coli, C.Jejuni, B.Cereus and A.hydrophila from examined chicken meat, chicken nuggets, chicken paneeh and chicken luncheon (n=25).

Samples	Salmonella		E.coli		C. jejuni		B. cereus		A. hydrophila	
	No	%	No	%	No	%	No	%	No	%
Chicken	14	56	17	68	19	76	15	60	7	28
meat										
Chicken	2	8	3	12	0	0	2	8	0	0
nuggets										
Chicken	3	12	3	12	4	16	0	0	1	4
paneeh										
Chicken	2	8	2	8	0	0	10	40	0	0
luncheon										

RESULTS

E.coli= Enteropathogenic Escherichia coli

B.C= Bacillus cereus

DISCUSSION

The muscle tissue and body fluids of healthy living animals are usually free from pathogenic bacteria, but during slaughtering and processing contamination occurs leading to introduction of pathogens into the meat. The source of these pathogens may be endogenous from the gastrointestinal tract or from surrounding environment in farm and/ or slaughterhouse. Poultry are the most common food vehicle of human infection with enteropathogens throughout the world.

Enteropathogens include:

A. Salmonella:

The results obtained in table (1) shows that the Salmonellae isolated from chicken meat, chicken nuggets, chicken paneeh and chicken luncheon at an incidences of 56 %, 8%, 12 % and 8% respectively. C.jejuni= Campylobacter jejuni A.hydrophila= Aeromonas hydrophila

The isolated Salmonella in chicken meat may be attributed to contamination during slaughtering and |or processing from workers' hands which nearly similar to results achieved by **Carraminana et al., (1997)** who stated that the prevalence of Salmonella in environmental samples ranged from 30% in feces to 75% in scald water samples. The Salmonella organisms found on carcasses at the post-spray wash site.

Nearly similar results were reported by **Wilson et al., (1996).** In contrary to our results **Uyttendale et al., (1999)** recorded that the samples which were frozen and kept at – 20 °C before analysis may have influenced the number of Salmonella recovered.

B- Escherichia coli:

E.coli was isolated with incidence of 68 %, 12 %, 12%, and 8% in chicken meat, chicken nuggets, chicken paneeh and

chicken luncheon, respectively. The highest incidence was in chicken meat which reveals that the high source of E.coli in the intestinal tract of the chicken which agree with **James et al.**, (1992) who found that the average log of E.coli count in poultry carcasses was 2.04 before chilling and 1.2 after chilling. On the other hand, the incidence

of E.coli was the lowest in chicken luncheon due to heat treatment.

The incidence of E.coli in chicken nuggets and chicken paneeh was lowered due to the effect of freezing which minimize the total count of E.coli, this in-accordance to **James et al.**, (1992) who found that the average log of E.coli count in poultry carcasses was 2.04 before chilling and 1.2 after chilling. On the other hand, the incidence of E.coli was the lowest in chicken luncheon due to heat treatment.

Generally the presence of E.coli in examined chicken meat and chicken meat products considered as an indicator for improper handling or unhygienic conditions which agreed with **Frazier and Westhoff (1983) and Hashim, (2003).** In contrary to **Abd EI -Haffeiz (1999)** who reported that the E.coli could not be detect from nuggets.

The most E.coli infection had been caused by serotype O157:H7 however, other serotypes had been identified and associated with outbreaks (Thomas et al., 1993). The transmission of *E.coli O157:H7* occurred primarily through the consumption of certain foods of particular importance to the chicken meat industry (Chapman et al., 1993).

C. Campylobacter jejuni:

Campylobacter jejuni was isolated from chicken meat and chicken paneeh, with incidence of 76 % and 16%, respectively. While could not isolated from chicken nuggets and chicken luncheon. These results indicated that chicken meat and chicken paneeh were contaminated with Campylobacter jejuni from intestinal content during slaughtering, defeathering and evisceration (**Adams and Moss, 2000**) or due cross contamination during handling and processing.

D. Bacillus cereus:

Bacillus cereus isolated from chicken meat, chicken nuggets and chicken luncheon at an incidence of 60 %, 8% and 40 % respectively and could not be detected in chicken paneeh.

These results attributed to contamination during slaughtering and defeathering of chicken especially dirty chicken. While, in chicken nuggets and chicken luncheon may be due to use of contaminated additives, seasonings and spices with Bacillus cereus spores. These results in-accordance with Konuma et al., (1988) and Kramer and Gilbert (1989).

E. Aeromonas hydrophila:

Aeromonas hydrophila only isolated from chicken meat and chicken paneeh at incidence of 28 % and 4 %, respectively. And could not be isolated from chicken nuggets and chicken luncheon. From these data we can conclude that, the contamination of meat and paneeh mainly from the water either used for drinking of chicken before slaughtering and / or water used for washing the carcass after evisceration where Aeromonas hydrophila could not be detected in chicken nuggets and chicken luncheon. These results agreed with (Biair et al., 2000) who reported that Aeromonas hydrophila has been frequently found in the red meat and poultry.

REFERENCES

Abdel-Haffeiz, E.M. (1999): Application of HACCP system in chicken nuggets to produce safety and high quality products. Alex. J. Vet. Sci. Vol 115 No 4.

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APHA, American Public Health Association. (1992): Compendium of Methods for the Microbiological Examination of Foods. 1015 Fifteenth Street, NW, Washington, DC 2005.

Adams, M.R. and Moss, M.O. (2000): Food microbiology 2nd Ed. PP 199 computape (Pickering) Ltd., North Yorkshire, England.

Aruno, O.O; Aydn, A.; Vural, A.I.; Cffcoglu, G. and Aksu, H. (2007): Determination of *E.coli O157* in raw and cooked Doner Kabab by using IMS technique. Medcyna-Weterynaryina, 63(10):1181-1183.

Biair, I.S.; McMahan, M. A.S. and McDowell, D.A. (2000): *Aeromonas*. In Encyclopedia of Food Science and Food technology and Nutrition. Harcourt Brace Jouanovich Publichers, New York, PP: 135 -170.

Carraminana, J.J.; Yanguela, J.; Blanco, D.; Rota, C.; Agustin, A.I.: Arino, A. and Herrera, A. (1997): *Salmonella* incidence and distribution of serotypes throughout processing in a Spanish poultry slaughterhouses. J. Food Prot., 60(11): 1312-1317.

Chapman, P. A.; Siddons, C.A.; Wright, D. J.; Norman, P.; Fox, J. and Crick, E. (1993): Cattle as a possible source of verocytotoxin-producing E.coli O157 infection in man. Epidemiol. Infect. 111(3): 439-447.

Cabedo, O.L.; Picart-i-Barrot, L. and Teixdoi-Canelles, A. (2008): Prevalence of Listeria Monocytogenes and Salmonella in ready-to-eat food in Catalonia, Spain. J. Food Prot., 71(4): 855-859.

Corry, J.E. L.; Mansfield, L. P.; Forsyth, S. J. and Ataby, H. I. ;(2001): Culture media for isolation of Campylobacter, Aerobacter and Helicobacter. In: Culture Media for Food Microbiology. 2nd Ed. (Edited by Corry. J. E. L.; Curtis, G.D. W. and Baird, R.M.) Amsterdam. Frazier, W. C. and Westhoff, D. C. (1983): Food microbiology. 3rd Ed., Tata McGraw Hill Publ. Co. New Delhi.

Granum, P. E. (2001): "*Bacillus cereus*" In, Food Microbiology: Fundamentals and Frontiers, 2nd Ed. ASM Press, Washington, D.C. P: 373- 381.

Hashim, E. S. Y. (2003): Aerobic and anaerobic enterotoxigenic bacteria in ready-to-eat food. Ph. D. Thesis, Fac. Vet. Med. Moshtohor, Zagazig Univ-Benha Branch.

ICMSF, International Commission on Microbiological Specification for Foods (1978): Microorganisms Ecology of food. University of Toronto Press. Vol 1, Toronto, Ontario, Canada.

James, W.O.; Brewer, R.L.; Prucha, J. C.; William, W.O. and Parharm, D.R. (1992): Effect of chlorination of chill water on the bacteriological profile of raw chicken carcasses and giblets. J. Amer. Vet. Med. Assoc., 200(1): 60-63.

Konuma, H.; Shingawa, K.; Tokumaru, M.; Onoue, Y.; Konno, S.; Fujino, N.; Shigegisa, T.; Kurata, H.; Kuwabara, Y. and Carlos, A. M. Lopes. (1988): Occurrence of *Bacillus cereus* in meat products, raw meat and meat product additives, J. Food Prot., 51(4): 324-326.

Kramer, J. M. and Gilbert, R. J. (1989): *Bacillus cereus* and other Bacillus species, P: 21-70. In M.P. Doyle (ed), Food-borne Bacterial pathogens. Marcel Dekker, New York, N.Y.

Lee, A.; Smith, S.C. and Coloe, P.J. (1998): Survival and growth of *Campylobacter jejuni* after artificial inoculation onto chicken skin as a function of temperature and packaging conditions. J. Food Prot., 61: 1609-1614.

Shriver-lake, L.C.; Turner, S.and Tailt, C.R. (2007): Rapid detection of *Escherichia coli O157:H7* spiked into food matrices. Anal. Chim. Acta; 584(1): 66-71.

Thomas, A.H.; Chart, T.; Cheasty, H.R.; Smith, J.A and Rowe, B. (1993): Verocytotoxin producing *Escherichia coli*, particularly serogroup O157 associated with human infection in the United Kingdom. Epidemiol. Infect. 110 (3): 591-600. Uyttendale, M.; DeTroy, P. and Debevere, J. (1999): Incidence of *Salmonella, Campylobacter jejuni, Campylobacter coli and Listeria monocytogens* in poultry carcasses and different types of poultry products for sale on the Belgian retail market. J. Food Prot., 62(7): 735-740.