



A Review: Herbal Medicine as an Effective Therapeutic Approach for Treating Digestive Disorders in Small Ruminants

Mohamed Zeineldin^{1*}, Mohamed Abdelmegeid², Radwa Barakat³, Mohamed Ghanem¹

¹ Department of Animal Medicine, College of Veterinary Medicine, Benha University, Egypt.

² Department of Animal Medicine, College of Veterinary Medicine, Kafr-Elsheikh University, Egypt.

³ Department of Comparative Biosciences, College of Veterinary Medicine, University of Illinois at Urbana-Champaign, USA.

ABSTRACT

Gastrointestinal (GIT) disorder is a significant health and economic problem that account for extraordinary financial loss in small ruminants. Despite the great efforts in management, GIT disorder remains the main cause of animal mortality, weight gain retardation and production loss in the small ruminant industry. Traditionally, herbal medicine has been used to treat and prevent various ruminant disorders. Herbs have nutritional and pharmaceutical properties that interact with one another polyvalently to increase the depth and breadth of the clinical effects compared with those seen in conventional pharmacological drugs. Recently, there has been an increase in the utilization of herbal medicine, reflecting an expanding trust in such therapeutic approaches. These methods have the advantage of utilizing locally available plants with high medicinal properties. Additionally, medicinal plants have the advantages of availability, compatibility with the body, simplicity of storage and negligible side effects. Here, we review representative examples of research available on the most commonly used traditional herbal remedies for the treatment of the most common digestive disorders (diarrhea, rumen insufficiency and GIT nematodes) in small ruminants. We also provide an extensive background on the recent advancement in the adoption of modern platforms in this field and research possibilities for future drug discovery and development.

Key words:

Gastrointestinal; herbal medicine; therapy; small ruminant.

*Correspondence to:

zeineldn@illinois.edu

1-INTRODUCTION

Small ruminants are a very important ecological niche among livestock, as they have a high digestion coefficient for converting poor feedstuffs to very useful products for humans, such as meat, milk and wool (Abd El-Raof et al., 2007). The economic significance of small ruminants in developing countries and the tropics cannot be overestimated (Ghanem et al., 2006). Subsequently, it is important to focus on the illnesses and disorders that influence small ruminants, with a specific end goal of improving and maintaining the profitability of these animals to benefit the human population in such countries.

Gastrointestinal disorders have been the main cause of animal mortality, weight gain retardation and production loss in small ruminant husbandry (Kaplan and Pesca, 2007; Zein Eldin et al., 2013). Varied efficacies of conventional medicaments against GIT disease have been

reported. However, concurrently, the pernicious effects of the overuse and misuse of these chemicals, the increased prevalence of resistance and high treatment cost necessitate the development of alternative therapies for the treatment and management of GIT diseases in small ruminants (Rahmann and Seip, 2007). These alternative treatment options include genetic resistance control, pasture management, nutrition adjustment, biological regulation, vaccination, and the use of herbal medicine (LeJambre et al., 2008). Recently, a large number of pharmaceutical companies and researchers from various fields have focused on the potential effectiveness of medicinal herbs as a wealthy resource for drug discovery (Malla et al., 2015; Hossen et al., 2016). In developing countries, approximately 25% of current therapies are developed from traditionally used medicinal plant a source, which has led to the discovery of 75% of the currently available herbal drugs (Jia and Tang, 2003). Meanwhile, in growing countries, the

contribution of medicinal plants to therapeutic medications is as high as 80% (Joy et al., 2001; Jia and Tang, 2003). The history of using herbal medicine to treat illness and enhance overall health has been reviewed by (Engel, 2007), with an emphasis on GIT diseases in small ruminants. Medicinal herbs have been proven to be economical, efficient, easily available and safe to use with almost no side effects (Hossen et al., 2016).

In this review, we focus on the previous and current status of traditional herbal medicine used for the treatment of selected GIT disorders (diarrhea, rumen insufficiency and GIT nematodes) in small ruminants. We also provide an extensive background on the recent advancement in the utilization of modern technology platforms in this field and possibilities for further research in the future.

2-Complexity of herbal medicine

Herbal medicine is generally defined as a pharmaceutical product extracted from the entire plant or from any part of a plant, including leaves, bark, roots, seeds and flowers (Winslow and Kroll, 1998). Herbal remedies may contain a single plant or a mixture of pharmacologically dynamic plants that are accepted as having synergetic impacts (Rotblatt and Ziment, 2002). Herbal plants are sold as either raw herbs or concentrates of the active substance from the entire plant or from the predefined part of the plant. Both the crude plant and the concentrate contain blends of organic chemicals, which may incorporate fatty acids, sterols, alkaloids, flavonoids, glycosides, saponins, tannins, and terpenes (Rotblatt and Ziment, 2002). In this manner, it is difficult to decide which constituent of the plant is responsible for the biological and pharmacological action. Moreover, the procedure for extracting the dynamic substance may change the concoction structure and the pharmacological action of the natural constituents. Additionally, a few natural variables (soil, distance, seasonal variability in temperature, humidity, daylight period, rainfall, shade, dew, and frost conditions) may influence the concentrations of the active plant compounds. Other vital variables, including seeding, harvesting time, genetic component, infections, and competition with other plant species, play a vital role in creating uniform herbal products (Wijesekera, 1991). Due to numerous elements that influence the congregation of active ingredients in the final products, producers have endeavored to make a more consistent product by recognizing certain interesting synthetic segments of a plant that are defined as markers and adjusting the production process to produce a reliable level of these markers in the final batch of

the herbal product. Currently, plant markers are defined to have a pharmacological spectrum, but very few are known to have clinical impacts (Zein-Eldin et al., 2014). Additionally, some producers have demonstrated that the standardization during the extraction process by adding purified active markers to the extract impacts the original balance of the organic ingredients in the final product (Ernst, 2005). Therefore, the toxicity and safety profile of the herbal extract should be considered before finalizing herbal products. Despite the multi-faceted quality of herbal products, their efficacy and desirability are superior to those of synthetic chemical drugs (Ernst, 2005).

3-Current technology in the herbal medicine industry

Recently, the international community has turned the modernization of the herbal medicine industry into a crucial component of the comprehensive acknowledgment of alternative therapy. The standardization and quality management of herbal treatment using current science and innovation have become critical. Herbal plants contain multiple active constituents, which can work harmoniously within the host body and yield particular therapeutic activities with minimal or no undesirable side effects. Therefore, the development of modern technology for the herbal therapy industry is likely to play a crucial role in providing high-quality herbal products to customers worldwide (Huie, 2002). For example, the use of modern technology in plant preparation can aid in the extraction of the required chemical ingredients from plants for further separation and characterization. These modern techniques include solid-phase micro-extraction, ultrasonic extraction, supercritical fluid extraction, pressurized liquid extraction, microwave-assisted extraction, solid-phase extraction, and surfactant-mediated extraction. A detailed explanation of the basic principles of modern sample-preparation techniques is available in many published reviews (Huie, 2002). After plant processing, identification of the concentration of biologically active components in the plant should be performed to achieve qualitative and quantitative estimates of quality based on sensitive analytical techniques (Porfírio et al., 2016). Another field of modern technology in herbal medicine is toxicogenomics (Borner et al., 2011), which targets the interaction between the structure and activity of the genome and the negative biological effects of exogenous agents. This field is dependent on the notion that the toxic impacts of xenobiotics on biological systems are generally reflected at the cellular level by their impact on gene expression (transcriptomics), protein determination

(proteomics) and the production of small metabolites (metabonomics). All of these techniques can be powerful tools for future study that will facilitate the development of highly sensitive and real-time detection and the evaluation of active substances in medicinal plants.

4-Herbal medicine use for the treatment of diarrhea in small ruminants

Diarrhea is one of the most significant economic problems in small ruminants and can cause great losses, especially in sheep herds due to the death of many affected animals, particularly in early life (Scott, 2007; Schoenian, 2007). The economic losses arise not only from lamb mortalities but also from the treatment cost and labor required to treat the sick animals. Moreover, acute diarrheal infections encountered in a herd are often difficult to control because of the large numbers of potential enteropathogens and the variability in individual animal immunity within the herd (Benavides et al., 2015). Diarrhea is not generally defined as a GIT disease but rather is a sign of other health problems in food-producing animals (Offiah et al., 2011). The etiology of diarrhea is complex and includes infectious agents, poor management, reproductive circumstances, dietary changes, and host immune status (Diaz-Lee et al., 2011). Thus, diarrhea continues to be the major therapeutic threat in small ruminant production systems (Grayson, 2011). In ruminants, different antimicrobials (oxytetracycline, neomycin, ampicillin, amoxicillin and colistin) are commonly administered for the treatment and control of diarrhea (Bywater, 2005). The use of antibacterial drugs orally, parentally, or by both routes simultaneously for the treatment of bacterial diarrhea is a controversial subject in both human and veterinary medicine due to the development of antimicrobial resistance (Suter et al., 2005). Antimicrobial treatment could additionally aggravate the GIT microbial communities and delay the clinical evolution of the diseased animals (Jiménez et al., 2007). Recently, the therapeutic use of medicinal herbs in the treatment of diarrhea has become a common practice due to the adoption of traditional folk medicines (Devi et al., 2013; Li et al., 2015). Medicinal plants contain multiple active ingredients, such as alkaloids, glycosides, flavonoids, terpenes and tannins, that have potential enhancing and/or neutralizing effects and are considered to be relatively safe for prolonged use (Maikai et al., 2010; Offiah et al., 2011; Gilani and Attaur, 2005). The active constituents of medicinal plants may alleviate diarrhea through interaction with several types of cells in the GIT tract, including epithelial cells, immune cells, commensal

microbiota, and pathogenic microbial communities (Teke et al., 2007). A wide range of herbs, including *Anacardium occidentale*, *Psidium guajava* (Lans and Brown, 1998), *Myristica fragrans* (Zain Eldin et al., 2013) and a granule mixture of *Coptischinensi*, *Magnolia officinalis*, *Atractylodes lancea*, *Prunus mume* and *Poria cocos* (Li et al., 2015), have been used for the treatment of diarrhea in small ruminants. In the study by Offiah et al. (2011), 132 medicinal herbs were examined through 248 questionnaires survey, in which 57 plants were properly identified for their antidiarrheal properties. The predominant plant families in this group are the Fabaceae (21%) and Combretaceae (14.04%) families. The medicinal properties of various parts of these herbs were also detected, with the leaves being the most commonly identified part. The route of administration mainly depended on the type of plant processing. Medicinal plants are usually processed by moistening the fresh or raw plant in water and used by drenching the whole extract. In some cases, the plant materials are blended into the diet to improve their palatability. In another study, 43 medicinal herbs belonging to 26 families were found to be used in ethnoveterinary practices for the treatment of GIT disorders in a livestock-rearing system (Mussarat et al., 2014). The results of this study demonstrated that *Zingiber officinale*, *Punica granatum*, *Triticum aestivum*, *Gossypium hirsutum*, and *Withania coagulans* were the most commonly approved species for diarrhea treatment. Medicinal plants used in the treatment of diarrhea in small ruminants are shown in Table 1. Generally, the herbs used in the treatment of diarrhea in small ruminants are administered orally to the affected animal; for example, the crushed stem of *Allium sativum* is boiled in water and decocted to treat gastrointestinal problems in goats and sheep (Casella et al., 2013). Phytochemical screening of *Allium sativum* confirmed the presence of allicin, which is responsible for its action against the gastrointestinal pathogens *Escherichia coli* and *Salmonella typhi* (Saravanan et al., 2010). Moreover, *Cassia fistula* fruit and the fruit cover of *Punica granatum* exhibit strong anti-*Escherichia coli* activity that may be due to the existence of various classes of phenols, flavonoids, terpenoids, saponins, steroids, anthraquinone, and glycoside compounds in the tested extracts (Bhalodia et al., 2012). Some plants are used in a mixture with other plants or as additives for herbal formulations; for example, the seeds of *Foeniculum vulgare* are mixed with different additives, and the root of *Glycyrrhiza glabra* is mixed with flour and used for GIT problems in ruminants. Manonmani et al. (2011) studied *Foeniculum vulgare* seeds in vitro and found that aqueous extracts of the seeds exhibit

strong inhibitory activity against gastrointestinal pathogens such as *Escherichia coli*, *Salmonella typhi*, and *Bacillus cereus*. The pharmacological justification of herbal remedies through different sampling procedures and extraction protocols should ensure strong validity and reliability of traditional knowledge of the respondents before any decision is made to use the plants for treatment (Devi et al., 2013). Therefore, further investigations of all the plants listed above should be performed to validate their efficacy in the treatment of diarrhea in small ruminants.

5-Herbal medicine for the treatment of rumen function insufficiency in small ruminants

In ruminant production systems, consuming high-concentrate feeds can impact rumen fermentation dysfunctions and cause serious GIT disorders such as lactic acidosis and bloating (Beauchemin and

Buchanan-Smith, 1990). Lactic acidosis is mainly defined as an acute GIT disorder (Zein-Eldin et al., 2014) and is clinically characterized by depression, anorexia, inactivity, abdominal distension, diarrhea and general weakness (Ullah et al., 2013). Lactic acidosis occurs due to a disproportionate consumption of readily fermentable carbohydrates that decrease ruminal pH to improper levels and lead to the rapid production of lactic acid with rapid fermentation (Mickdam et al., 2016). Lactic acidosis is considered a dramatic form of ruminal microbial fermentative disorder and is lethal in less than 24 hours in some cases (Huo et al., 2014). In addition to lactic acidosis, sheep are subjected to many other GIT diseases; one of these important diseases is bloat, which is a costly disease due to the grazing of these sheep on high protein pastures (Stanford et al., 2001).

Table 1. Medicinal plants used in the treatment of diarrhea in small ruminant

Botanical name	Family	Used part	Citation
<i>Adansonia digitata</i>	Bombacaceae	Fruits	Offiah et al., 2011
<i>Albizia amara</i>	Fabaceae	Bark	Maroyi, 2012
<i>Aloe greatheadii</i>	Asphodelaceae	Leaves	Maroyi, 2012
<i>Anacardium occidentale</i>	Anacardiaceae	Bark	Lans and Brown, 1998
<i>Atractylodes lancea</i>	Asteraceae	Roots	Li et al., 2015
<i>Capsicum annum</i>	Solanaceae	Fruit	Maroyi, 2012
<i>Cassia fistula</i>	Caesalpinaceae	Whole plant	Mussarat et al., 2014
<i>Cinnamomum zeylanicum</i>	Lauraceae	Inner bark	Lans et al., 2007
<i>Citrus reticulata</i>	Rutaceae	Leaves	Mussarat et al., 2014
<i>Daniellia oliveri</i>	Fabaceae	Bark	Chah et al., 2009
<i>Dioscorea dametorum</i>	Dioscoreaceae	Root	Chah et al., 2009
<i>Ficus burkei</i>	Moraceae	Root	Maroyi, 2012
<i>Gossypium hirsutum</i>	Malvaceae	Biennial	Mussarat et al., 2014
<i>Khaya senegalensis</i>	Meliaceae	Leaves	Offiah et al., 2011
<i>Landolphia spp</i>	Apocynaceae	Leaves	Chah et al., 2009
<i>Lepidium sativum</i>	Brassicaceae	Seed	Araya et al., 2015
<i>Leucaena spp</i>	Fabaceae	Leaves	Chah et al., 2009
<i>Magnolia officinalis</i>	Magnoliaceae	Flowers	Li et al., 2015
<i>Manihot esculenta</i>	Euphorbiaceae	Roots	Chah et al., 2009
<i>Meriandra dianthera</i>	Lamiaceae	Leaf	Araya et al., 2015
<i>Musa paradisiacal</i>	Musaceae	Fruit	Mirani et al., 2014
<i>Myristica fragrans</i>	Myristicaceae	Seed	Zein Eldin et al., 2013
<i>Myrothamnus flabellifolius</i>	Myrothamnaceae	Root	Maroyi, 2012
<i>Oryza sativa</i>	Poaceae	Seed	Mirani et al., 2014
<i>Parinari curatellifolia</i>	Chrysobalanaceae	Bark	Maroyi, 2012
<i>Plantago lanceolata</i>	Plantaginaceae	Leaves and Husk	Mirani et al., 2014
<i>Poria cocos</i>	Polyporaceae	filaments	Li et al., 2015
<i>Potentilla recta</i>	Rosaceae	Aerial parts	Lans et al., 2007
<i>Prosopis cineraria</i>	Fabaceae	Leaves	Mirani et al., 2014
<i>Prunus mume</i>	Rosaceae	Flower	Li et al., 2015
<i>Psidium guajava</i>	Myrtaceae	Buds, leaves	Lans and Brown, 1998
<i>Punica granatum</i>	Lythraceae	Fruits	Mussarat et al., 2014
<i>Salix alba</i>	Salicaceae	Bark	Lans et al., 2007
<i>Sarcostemma viminalis</i>	Asphodelaceae	Stem	Maroyi, 2012
<i>Senna sp.</i>	Fabaceae	Pod	Lans et al., 2007
<i>Solanum spp</i>	Solanaceae	Leaves	Chah et al., 2009
<i>Triticum aestivum</i>	Poaceae	Stem	Mussarat et al., 2014
<i>Ulmus fulva</i>	Ulmaceae	Inner bark	Lans et al., 2007
<i>Vernonia conferta</i>	Asteraceae	Leaves	Chah et al., 2009
<i>Withania coagulans</i>	Solanaceae	Leaves /Fruits	Mussarat et al., 2014
<i>Xeroderris stuhlmannii</i>	Fabaceae	Bark	Maroyi, 2012
<i>Zea mays</i>	Poaceae	Grains	Chah et al., 2009
<i>Zingiber officinale</i>	Zingiberaceae	Rhizome	Mussarat et al., 2014

Table 2. The most common species of gastrointestinal nematodes infecting small ruminants

Gastrointestinal nematodes species	Location in the host
<i>Haemonchus contortus</i>	Abomasum
<i>Teladorsagia circumcincta</i>	Abomasum
<i>Trichostrongylus axei</i>	Abomasum
<i>Trichostrongylus colubriformis</i>	Anterior small intestine
<i>Trichostrongylus vitrinus</i>	Anterior small intestine
<i>Trichostrongylus rugatus</i>	Small intestine
<i>Cooperia curticei</i>	Small intestine
<i>Nematodirus spathiger</i>	Small intestine
<i>Nematodirus filicollis</i>	Small intestine
<i>Bunostomum</i>	Small intestine
<i>trigonocephalum</i>	
<i>Oesophagostomum</i>	Large intestine
<i>columbianum</i>	
<i>Oesophagostomum venulosum</i>	Large intestine
<i>Chabertia ovina</i>	Large intestine

Bloat is mainly caused by the failure of the animal to release gas pressure (Stanford et al., 2001). Treatment of such conditions mainly relies on prohibiting concentrates and feeding hay to stimulate saliva secretion. Additional lactic acidosis treatment includes oral antacids, such as sodium bicarbonate and oral electrolyte solutions, and the administration of antimicrobial treatments, such as penicillin, potentiated sulphonamides and tetracycline (Baruah, 2008). Antimicrobial treatment should be given with other supportive treatments to control the adverse effects of histamine release and endotoxemia.

Current scientific research highlights the potential significance of using plant-based medicine to enhance rumen function in ruminants. However, many of these studies are only laboratory-based. Nevertheless, their results provide evidence that plant extracts and their constituents have the potential to enhance rumen function and feed utilization in ruminants (Benchaa et al., 2008). In livestock husbandry systems, antibiotics are widely administered to prevent infection and improve feed utilization. The emergence of antibiotic-resistant bacteria has increased public health concerns regarding the conventional use of antimicrobial drugs in livestock feed. Consequently, a considerable attempt has been made to develop alternatives to antimicrobial therapies. Several theories have been proposed to explain the mechanism of the antibacterial action of plant extracts. It is believed that the active ingredients in the plant extract exert their activities by interaction with a wide variety of cellular processes associated with the bacterial cell membrane, including electron transport, ion gradients, protein translocation, phosphorylation, and other enzyme-dependent reactions (Dorman and Deans, 2000; Acamovic and Brooker, 2005).

Active areas of plant metabolite research have demonstrated that some medicinal plants,

especially those containing essential oils, flavonoids, saponin and tannin, enhance ruminal fermentation and improve nutrient utilization in ruminants (Hristov et al., 1999; Rochfort et al., 2008). The inclusion of some plant extracts in ruminant feed results in the inhibition of deamination and methanogenesis processes, a reduction of acetate or methane production, and an increase in propionate and butyrate concentrations (Calsamiglia et al., 2007). For this reason, ruminant nutritionists have become interested in using natural products instead of chemical compounds as manipulators of rumen function to improve feed efficiency and increase ruminant productivity (Santoso et al., 2006).

Recently, anise oil, garlic oil, cinnamaldehyde, gentian, oregano, eugenol, and cloves have been studied as an alternative to antimicrobials and as a natural alternative growth promoter in livestock (Ertas et al., 2005). Plant extracts containing essential oils have the potential to inhibit the development of induced lactic acidosis using rumen fluid as an in vitro batch system (Hutton et al., 2009). However, knowledge of the underlying mechanism and an explanation of this protective effect are lacking. While some herbs were able to maintain pH, and inhibit both overall microbial activity and gas production, others appeared to have more extensive effects. The use of such natural plants is under explored in sheep. In contrast, the effects and mechanisms of the action of plants containing saponins and tannins on rumen microbial fermentation have been extensively researched and reviewed (Piacente et al., 2005). Plants containing a range of proanthocyanidins also have considerable effects on ruminal microbial population and the inhibition of foam production (Sivakumaran et al., 2004). In the study of Zeineldin et al. (2014), gentian root was used as an alternative to the ordinary treatment of induced lactic acidosis in sheep, and this experimental study

proved that this herb could be used to modulate and restore rumen function. From the previous literature, considerable awareness has been raised on the potential use of herbal medicine to modulate rumen microbial fermentation. However, several controversies need to be resolved before specific recommendations for commercial use can be made. Future research into additional medicinal plants that could be of interest is needed.

6- Herbal medicine as an alternative treatment for gastrointestinal nematodes

GIT parasitism remains one of the major health and economic problems in small ruminant husbandry worldwide, particularly in ruminants owned by the poor populations (Canul-Ku et al., 2012; Carvalho *et al.*, 2012). The most common gastrointestinal parasite species found in small ruminants are listed in Table 2. Anthelmintic treatment of GIT parasites is currently inexpensive and simple; however, as nematodes develop anthelmintic resistance, treatment will become less viable and the economic impact of GIT parasites will increase (Stear et al., 2007). Anthelmintic resistance is defined as a heritable change in the ability of individual parasites to survive the prescribed therapeutic doses of an anthelmintic drug (Prichard et al., 1980; Prichard, 1994; Coles, 2005). As a result of anthelmintic resistance, and the public health concern regarding the use of chemical therapeutics in the livestock industry, many research studies have been designed to examine alternative strategies for treating GIT parasites. The bioactive constituent produced by medicinal herbs to counteract GIT nematodes is of growing interest in anthelmintic drug research as a promising alternative to conventional anthelmintic drugs.

Until recently, most of the evidence of the anti-parasitic activities of herbal extracts and the determination of therapeutic doses have been preliminary and lacked scientific validity (Attindéhou et al., 2012). Various herbs have been used to treat parasitic diseases either through feeding ruminants the entire plant or administering plant extracts to the diseased animal (Jabbar et al., 2007), and several of them have been scientifically approved (Hordegen et al., 2003; Hasan et al., 2015; Macedo et al., 2015; Sanhokwe et al., 2016). For example, the seeds of squash and pumpkins, garlic, onion, walnuts, dill, parsley, mint and many other vine crops are accepted to contain a deworming substance and have been considered for use in sheep with varying outcomes (Strickland et al., 2009). In addition, ginger has been utilized as an anthelmintic plant for ruminants (Iqbal et al., 2006). The use of Acacia, Artemisia and tobacco extracts in parasitic animals has resulted in a decrease in the number of worms (Deharo et al., 2001). Throughout many

years of study, numerous herbs have been identified as having the potential to be used as deworming agents for livestock. Table 3 provides a representative list of these plants.

6-Conclusion and future direction

This review article aimed to cite the most common medicinal plants for the treatment of the most common GIT disorders in small ruminants and to document the interest among scientists in utilizing natural methods for the benefit of the livestock production industry. This therapeutic approach has the advantages of having fewer side effects and being cost effective, highly available and beneficial to the health of small ruminants. The present records of the medicinal uses of herbs may serve to deepen our knowledge and elucidate the mechanisms of action of these herbs in treating a range of small ruminant disorders. The rapid advancement of modern technology creates a good opportunity to employ such technologies in herbal therapy preparation and analysis to improve the quality of herbal products and understand the underlying mechanisms of action. Most of these technologies are not standardized, which will be the main challenge for pharmaceutical companies to overcome in establishing uniform interpretation methods. Considering the aforementioned issues, future studies are needed to reach a conclusion about the reliability, bioavailability, permeability and effective safe doses of active medicinal compounds for clinical trials.

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Conflict of interest

The authors declare no conflict of interest.

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amphistome *Gastrothylax indicus*. J. Parasit. Dis. 1-5.

Table 3. Medicinal plants used in the treatment of GIT nematodes in small ruminants.

Botanical name	Family	Used part	Citation
<i>Acokanthera oppositifolia</i>	Apocynaceae	Shrub	Sanhokwe et al., 2016
<i>Adhatoda vesica</i>	Acanthaceae	Root and leaves	Khan et al., 2012
<i>Albizia anthelmintica</i>	Fabaceae	Bark	Grade' et al., 2008
<i>Allium cepa</i>	Amaryllidaceae	Bulbs	Mehlhorn et al., 2011
<i>Allium sativum</i>	Liliaceae	Bulbs	Hasan et al., 2015
<i>Aloe ferox</i>	Asphodelaceae	Flowers	Sanhokwe et al., 2016
<i>Ananas comosus</i>	Bromeliaceae	Leaves	Hordegen et al., 2003
<i>Ananas comosus</i>	Bromeliaceae	Leaves	Shaha et al., 2015
<i>Artemisia absinthium</i>	Asteraceae	Aerial parts	Tariq et al., 2009
<i>Artemisia campestris</i>	Asteraceae	Leaves	Akkari et al., 2014
<i>Artemisia maritima</i>	Asteraceae	Whole plant	Irum et al., 2015
<i>Artemisia vestita</i>	Asteraceae	Whole plant	Irum et al., 2015
<i>Azadirachta indica</i>	Meliaceae	Leaves and Seeds	Hördegen et al., 2003; Shaha et al., 2015
<i>Butea monosperma</i>	Fabaceae	Seeds	Singh et al., 2015
<i>Caesalpinia crista</i>	Caesalpiniaceae	Seeds	Hördegen et al., 2003; Mushtaq et al., 2015
<i>Calliandra portoricensis</i>	Leguminosae	Roots, leaves, flowers	Iqbal et al., 2005
<i>Calotropis (C.) procera</i>	Asclepiadaceae	Flowers	Iqbal et al., 2005
<i>Calotropis procera</i>	Apocynaceae	Leaves and flowers	Aggarwal et al., 2015
<i>Carica papaya</i>	Caricaceae	Leaves and seeds	Attindehou et al., 2012
<i>Carissa caranta</i>	Apocynaceae	Root	Khan et al., 2012
<i>Cassia obtusifolia</i>	Caesalpiniaceae	Leaves	Saidou et al., 2015
<i>Chenopodium album</i>	Chenopodiaceae	Whole plant	Jabbar et al., 2007
<i>Chenopodium ambrosioides</i>	Amaranthaceae	Leaves	Ketzs et al., 2002
<i>Chrysanthemum cinerariaefolium</i>	Asteraceae	Flowers	Mbaria et al., 1998
<i>Chrysophyllum cainito</i>	Sapotaceae	Stem	Fernandez, 1991
<i>Citrus sinensis</i>	Rutaceae	Peels	Gaínza et al., 2015
<i>Cocos nucifera</i>	Arecaceae	Fruit	Mehlhorn et al., 2011
<i>Cymbopogon citratus</i>	Poaceae	Leaves	Macedo et al., 2015
<i>Elephantorrhiza elephantina</i>	Fabaceae	Leaves	Sanhokwe et al., 2016
<i>Embelia ribes</i>	Myrsinaceae	Fruits	Hördegen et al., 2003
<i>Eucalyptus globulus</i>	Myrtaceae	Leaves	Kanojiya et al., 2015
<i>Eucalyptus staigeriana</i>	Myrtaceae	Leaves	Macedo et al., 2010
<i>Fumaria parviflora</i>	Fumariaceae	Whole plant and leaves	Hördegen et al., 2003; Mushtaq et al., 2015
<i>Hedera helix</i>	Araliaceae	Ripe fruits	Egual et al., 2007
<i>Hilderbrandtia sepalosa</i>	Myrtaceae	Roots	Gathuma et al., 2004
<i>Iris kashmiriana</i> Linn	Iridaceae	Dried plant parts	Khan et al., 2016
<i>Khaya senegalensis</i>	Meliaceae	Bark	Ademola et al., 2004
<i>Lansium domesticum</i>	Meliaceae	Whole plant	Kumarasingha et al., 2016
<i>Lepedeza cuneata</i>	Fabaceae	Leaves	Ahmed et al., 2014
<i>Leucaena leucocephala</i>	Fabaceae	Leaves	Hernandez et al., 2014
<i>Linariantha bicolor</i>	Acanthaceae	Roots and leaves	Kumarasingha et al., 2016
<i>Lippia sidoides</i>	Verbenaceae	Leaves	Camurc,a-Vasconcelos et al., 2008
<i>Maesa lanceolata</i>	Myrsinaceae	Leaves and fruits	Tadesse et al., 2009
<i>Melaleuca quinquenervia</i>	Myrtaceae	Leaves	Gaínza et al., 2015
<i>Melia azedarach</i>	Meliaceae	Fruits	Hördegen et al., 2003
<i>Moringa oleifera</i>	Moringaceae	Leaves	Attindehou et al., 2012
<i>Musa spp (banana plant)</i>	Musaceae	Leaves	Gregory et al., 2015
<i>Myrsine afriacana</i>	Primulaceae	Fruits, mixed with leaves	Gathuma et al., 2004
<i>Nauclea latifolia</i>	Rubiaceae	Bark	Onyeyili et al., 2001
<i>Nicotiana tabacum</i>	Solanaceae	Leaves	Iqbal et al., 2006
<i>Nigella sativa</i>	Ranunculaceae	Seeds	El-Far et al., 2014
<i>Ocimum sanctum</i>	Lamiaceae	Leaves	Kanojiya et al. 2015
<i>Pelargonium endlicherianum</i>	Geraniaceae	Roots	Kozan et al., 2016
<i>Peltophorum africanum</i>	Fabaceae	Root bark	Bizimenyera et al., 2008
<i>Petiveria alliacea</i>	Phytolaccaceae	Root	Lans and Brown, 1998
<i>Picria fel-terrae Lour</i>	Scrophulariaceae	Whole plant and leaves	Kumarasingha et al., 2016
<i>Piliostigma reticulatum</i>	Caesalpiniaceae	Leaves	Saidou et al., 2015
<i>Piptadenia viridiflora</i>	Fabaceae	Leaves	Morais-Costa et al., 2016
<i>Pistacia lentiscus</i>	Anacardiaceae	Shrub	Amit et al., 2013
<i>Plectranthus punctatus</i>	Lamiaceae	Aerial parts	Tadesse et al., 2009
<i>Punica granatum</i>	Punicaceae	Fruit peel	Aggarwal et al., 2015
<i>Ruellia tuberosa</i>	Acanthaceae	Root	Lans and Brown, 1998
<i>Salix babylonica</i>	Salicaceae	Leaves	Cedillo et al., 2015
<i>Sesbania grandiflora</i>	Fabaceae	Leaves	Azrul et al., 2016
<i>Spondias mombin</i>	Anacardiaceae	Leaves	Attindehou et al., 2012
<i>Thymus vulgaris</i>	Lamiaceae	Leaves	Ferreira et al., 2016
<i>Tinospora rumphii</i>	Menispermaceae	Stem	Fernandez, 1991
<i>Trichilia clausenii</i>	Meliaceae	Fruits	Cala et al., 2001
<i>Vernonia anthelmintica</i>	Asteraceae	Seeds	Hördegen et al., 2003
<i>Xanthoxylum zanthoxyloides</i>	Rutaceae	Leaves	Attindehou et al., 2012
<i>Zingiber officinale Roscoe</i>	Zingiberaceae	Rhizomes	El-Far et al., 2014;Matthews et al., 2016

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